# Performances of advanced bread wheat (*Triticum aestivum* L.) mutant lines for grain yield and some yield components

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#### Abstract

This research was conducted to assess yield and some yield components of 65 advanced bread wheat mutant lines of M<sub>6</sub> generation selected from M<sub>4</sub> populations of 4 different bread wheat genotypes. They were sown using an augmented design consisting of five blocks of which every block had 13 mutant advanced lines and 4 check genotypes (parents) in the 2014-2015 growing season. According to the data obtained from the experiment, there was a great variation for plant height (69.15-119.56 cm), spike length (9.26-11.78 cm), number of grains per spike (30.59-59.84), grain weight per spike (1.43-3.13 g), harvest index (32.57-48.37%), thousand grain weight (34.13-55.93 g) and grain yield (4712-9515 kg ha<sup>-1</sup>). The 33 lines for plant height, 22 lines for spike length, 21 lines for the number of grains per spike, 31 lines for grain weight per spike, 35 lines for harvest index, 30 lines for 1000 grain weight and 33 lines for grain yield have been found to perform better than check genotypes. According to the parents in more than one trait. These mutant lines were selected for pre-registration yield trials, and they may prove useful as a parent for yield improvement in a wheat breeding program. Gamma irradiations with 100 and 200 Gy may be more beneficial for yield improvement in wheat mutation breeding.

Keywords: Bread wheat; mutant line wheat; augmented; grain yield; yield components

#### INTRODUCTION

Mutagenesis and hybridization are generally the most commonly used breeding methods to develop new superior varieties. Mutation breeding has some advantages compared to cross breeding. When comparing mutation and hybridization; the occurrence of even a few desirable mutations in high yielding varieties has the great advantage of becoming homozygous and expressing its superiority within a couple of generations after induction in M<sub>2</sub> or M<sub>3</sub> as compared to  $F_6$  or  $F_7$  generations in case of hybridization (Chakraborty & Paul, 2013). Mutation breeding has become an appropriate option to improve plant characters when conventional breeding does not work, or the desired traits were recessive, or improving another character in an established plant variety, or improving one or two main character(s) (Ahloowalia & Maluszynski, 2001; van Harten, 1998). Besides, mutagenesis can isolate mutants with multiple characters, as compared to transgenes where the only line can be introduced it's the major advantage of inducing mutations (Louali et al., 2015). Mutation breeding can be applied to improve a specific character without changing other characters and it is possible to improve a single line without causing an important disturbance in the genome. The selection of tropical wheat mutant lines using selection index and multivariate analysis were considered effectively with a high determination value of 0.57 (57%). The results of the selection index show that seven lines were recommended for multi-location trials since they had a better selection index potential than the Dewata variety (Fadli et al., 2022). Furthermore, it may create a new character that did not belong to parental plants. The mutants developed in wheat have a great potential for direct release and inclusion in hybridization breeding programs (Sakin et al., 2005). The released mutant cultivars in different crops had a great economic impact on agriculture and food production and added billions of dollars to the economy of many countries (Jain, 2006). More than 3000 varieties of different crops have been officially released by mutation breeding techniques. Mutation induction with radiation has been the most frequently used method to develop direct mutant varieties (64% with gamma-rays, 22% with X-rays), accounting for about 90% of obtained varieties (Ahloowalia et al., 2004). Gamma-ray mutagen was effective in broadening genetic variability and increasing the means of wheat cultivars for grain yield and its components, helping plant breeders to practice an efficient selection in the M<sub>2</sub> and next mutated generations (Khanna et al., 1986; Al-Naggar et al., 2007).

The mutant lines showed the capacity to biofortify wheat grain without negatively impacting crop productivity and this population offers promising donors for improving grain parameters such as GA, length, width and quality. The data presented showed how the genetic variation generated through radiation could be used to test the linkage between various important grain parameters (Miller, 2017). Mutant populations have now been created for many cereal crops, including rice (Suzuki et al., 2008), durum wheat (Başer et al., 1997; Sakin & Yildirim, 2004) and bread wheat (Slade et al., 2005). Three seeds from each spike per plant of M<sub>1</sub> plants were collected from this M<sub>2</sub>, 17 desirable putative mutant plants which varied significantly with the mother were visually selected. Most of the mutant lines showed homogeneity for most of the characters studied. Eleven of these 17 lines were found to be promising in respect of days to flower, plant height (for semi-dwarf) and other traits including grain yield (Albokari, 2014).

The goal of a wheat breeding program is to develop superior genotypes as a result of many years of selection. Early generation selection is based on visual observations of yield components (spike length and size, number of spike per plant, number of spikelets per spike), disease resistance, tillering potential, lodging resistance and seed quality. Efficient evaluation of such large numbers of entries in field experiments is laborious, expensive, and dif-

ficult to manage because of the confounding effects that genotype x environment interaction and soil heterogeneity. In the early stage of the selection process, there could be an insufficient seed of the new treatments' to undertake replicated experiments or the number of genotypes could be very large to manage in terms of resources. Therefore, augmented designs have been developed for the evaluation of genotypes in the early stages of a breeding program (Federer, 1956). Augmented designs consist of two kinds of treatments, the checks or the standard treatments and new or augmented treatments. The design presumes checks as fixed effects whereas the new entries as random effects. The new entries are usually not replicated owing to a large number of entries initially in a breeding program, especially when dealing with large germplasm sets. However, the checks are replicated to act as points of reference. More sophisticated augmented designs allow for the adjustment of test varieties by rows and columns (Federer & Raghavarao, 1975; Lin & Poushinsky, 1983). The efficiency of the analysis over another is usually measured in terms of reduced error variance, expected error mean square or average standard error of the difference between genotype means (Binns, 1987; Cochran & Cox, 1957; Magnussen, 1990). The average standard error of the difference (SED) was reported to be more appropriate since it is used for comparison among genotypes using the same scale as the traits (Binns, 1987; Cullis & Gleeson, 1991; Gleeson, 1997). This study, it was aimed to determine the yield and yield components of M<sub>c</sub> advanced bread wheat mutant lines selected from M<sub>4</sub> populations induced different gamma rays of 4 different bread wheat genotypes.

# MATERIAL AND METHODS

*Experimental site and growing conditions:* This study was conducted at the University of Tekirdağ Namık Kemal, Faculty of Agriculture, Department of Field Crops, Turkey in the 2014-2015 growing season. Tekirdağ district locates at latitude 40° 36'-40° 31' and longitude 26° 43'-28° 08'and altitude is 10 m. The total precipitation was 435.1 mm and the average temperature was 11.5 °C during the 2014-2015 wheat growing season (November 2014-June, 2015) when the experiment was conducted. It is noted that the annual precipitation is about 30 mm

lower than the long term average (466 mm) and the average temperature is similar to the long term average (11.5  $^{\circ}$  C). According to soil analysis results, the experimental area's soil was clay-loam, slightly acidic (pH 6.5), limeless, and poor (1.08%) in the organic matter.

*Experimental materials, gamma irradiation and design:* Four bread wheat (*Triticum aestivum* L.) genotypes, Avusturalya, Bezostaja 1, Kate A-I and IBWSN4 were used as the parent material. The moisture contents of seeds of the genotypes were 12.1% for Avustralya, 11.4% for Bezostaja 1, 11.7% for Kate A-I and 12.0% for IBWSN4.

Gamma treatment was obtained from <sup>60</sup>Cobalt, Ob-Servo Sanguis Co-60 Research Irradiator with isotope model, while the dose rate was 2.190 kGy h<sup>-1</sup> before the 2009-2010 growing season sowing at the Turkish Atomic Energy Authority, Sarayköy Nuclear Research and Training Center, Ankara, Turkey. Right after irradiation, the experiment was set up using a total of 20  $M_0$  (100, 200, 300, 400 and 500 Gy) combination seeds together with the un-irradiated (control) in the experimental field of the Field Crops Department of the Faculty of Agriculture of Tekirdağ Namık Kemal University during the growing season of 2009-2010. The seeds obtained from the harvested plants in M<sub>1</sub> generation were sown in 2010-11 (M<sub>2</sub>), 2011-12 (M<sub>3</sub>) and 2012-13 (M<sub>4</sub>) growing seasons as 20 cm row distance in 5 meters of a 6-row plot with 4 replicate and as 400 seeds in each row. The experiment was carried out

in augmented design with 5 blocks with 65 advanced bread wheat mutant lines of M<sub>6</sub> generation selected from M<sub>4</sub> populations of 4 different bread wheat genotypes and their parents (Table 1) in the 2014-2015 growing season. Each block included 13 advanced bread wheat mutant lines and 4 check bread wheat genotypes (A, IB, K and B). Sown were made on Nov.7, 2014, by hand at the rate of 500 seeds per  $m^2$ and were 5 m in length, with 2 rows 0.2 m apart. Nitrogen and P<sub>2</sub>0<sub>5</sub> at 140 and 70 kg ha<sup>-1</sup>, respectively, were incorporated into the soil as compound fertilizer (20-20-0) before sowing, urea during tillering and ammonium nitrate before heading. The crop was kept free of weeds by hand hoeing when necessary. Morphological and yield characters were recorded on 15 random and guarded plants to study the effect of irradiation doses on the studied genotypes on plant height (PH- cm), spike length (SLcm), the number of grain per spike (NGS-no), grain weight per spike (GWS-g), thousand grain weight (TGW-g), harvest index (HI-%), and grain yield (GY) was calculated as kg ha<sup>-1</sup>.

The data set has been analyzed by the help of JUMP statistics package program in accordance with augmented experimental design was used. The characters averages that were statistically significant according to the variance analysis results have been compared to the Student-Newman-Keuls test ( $p \le 0.01$ ) and the alphabetical order of statistical significance groups obtained are shown in Tables 2 and 3 for each character (Gomez & Gomez, 1984).

Table 1. The experiment material of the study

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C1 (Bezostaja 1)	MT12 (400 Gy)	MT19 (200 Gy)	MT35 (400 Gy)	MT46 (300 Gy)	MT57 (200 Gy)			
MT1 (100 Gy)	MT13 (400 Gy)	MT20 (200 Gy)	MT36 (400 Gy)	MT47 (300 Gy)	MT58 (200 Gy)			
MT2 (100 Gy)	MT23 (500 Gy)	MT21 (200 Gy)	MT37 (400 Gy)	MT48 (300 Gy)	MT59 (200 Gy)			
MT3 (100 Gy)	MT24 (500 Gy)	MT22 (300 Gy)	MT38 (400 Gy)	MT49 (400 Gy)	MT60 (300 Gy)			
MT4 (100 Gy)	MT25 (500 Gy)	MT27 (300 Gy)	C3 (Kate A-1)	MT50 (400 Gy)	MT61 (300 Gy)			
MT5 (200 Gy)	MT26 (500 Gy)	MT28 (300 Gy)	MT39 (100 Gy)	MT51 (400 Gy)	MT62 (300 Gy)			
MT6 (200 Gy)	C2 (IBWSN 4)	MT29 (300 Gy)	MT40 (100 Gy)	MT52 (400 Gy)	MT63 (400 Gy)			
MT7 (200 Gy)	MT14 (100 Gy)	MT30 (300 Gy)	MT41 (100 Gy)	MT53 (400 Gy)	MT64 (400 Gy)			
MT8 (300 Gy)	MT15 (100 Gy)	MT31 (300 Gy)	MT42 (100 Gy)	C4 (Avustralya)	MT65 (400 Gy)			
MT9 (300 Gy)	MT16 (100 Gy)	MT32 (400 Gy)	MT43 (200 Gy)	MT54 (100 Gy)				
MT10 (300 Gy)	MT17 (100 Gy)	MT33 (400 Gy)	MT44 (200 Gy)	MT55 (100 Gy)				
MT11 (400 Gy)	MT18 (200 Gy)	MT34 (400 Gy)	MT45 (200 Gy)	MT56 (100 Gy)				
C: check genotype, MT: advanced mutant line								

# **RESULTS AND DISCUSSION**

As a result of the analysis of variance, the differences between the means of the mutant lines were found to be statistically significant at 0.01 probability level for plant height, spike length, thousandgrain weight, the number of grains per spike, grain weight per spike and harvest index and grain yield. The check (parent) mean performances, the adjusted mean performances of the mutant bread wheat lines calculated taking these into account and the results of the significances test are given in Table 2 and Table 3 for the investigated characters.

*Plant height (PH) (cm):* One of the most obvious and expected effects of mutagen applications on plants is the reduction in plant height. Considering this character, 65 advanced mutant lines averaged between 69.13-119.56 cm, while the averages of the checks varied between 92.86 - 117.74 cm (Table 2).

Among the check genotypes, the shortest and the longest plant height were measured in the IBWSN-4 line and the Australian variety respectively (Table 2). The twenty-three mutant lines gave a shorter plant height than the check genotype IBWSN-4, which has the shortest plant height. Among them, 13 lines were below the plant height value of 90.00 cm, which

is recommended for our region to avoid lodging problems. As a result, it is understood that these 13 advanced bread wheat mutant lines should be evaluated in the next generations in terms of plant height. Our results are supported by the findings of Shubhra et al. (2013), explaining that mutant lines with shorter plant lengths can be obtained as a result of gamma irradiation.

Spike length (SL) (cm): One of the most important yield components affecting wheat yield is the spike length and it is accepted as one of the most

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PH (cm)	Genotypes	SL (cm)	Genotypes	NGS (no)	Genotypes	GWS (g)
117.74 abc	C1	10.78 a-l	C1	42.98 e-x	C1	1.91 k-u
100.52 f-u	C2	10.46 b-t	C2	47.76 b-p	C2	2.12 g-t
92.86 n-A	C3	10.51 a-s	С3	48.56 a-o	C3	2.30 c-r
105.80 c-k	C4	10.35 b-u	C4	43.46 e-x	C4	2.11 g-t
119.56 a	<b>MT14</b>	11.78 a	MT61	59.84 a	<b>MT14</b>	3.13 a
118.96 ab	MT35	11.45 ab	MT44	59.09 abc	MT31	3.02 ab
117.66 a-d	MT31	11.38 abc	MT30	56.44 a-d	MT28	3.01 ab
117.66 a-d	MT48	11.24 a-d	MT31	54.74 а-е	MT33	2.92 abc
113.78 а-е	MT36	11.20 а-е	MT59	54.24 а-е	MT30	2.87 a-d
110.18 a-f	MT27	11.07 a-g	MT28	54.04 a-f	<b>MT27</b>	2.85 а-е
110.05 a-f	MT17	10.96 a-h	MT27	53.44 a-g	MT32	2.81 a-f
109.88 a-g	MT25	10.90 a-k	MT14	53.09 a-h	MT35	2.72 a-g
109.88 a-g	MT40	10.86 a-l	MT42	52.19 a-1	MT37	2.71 a-g
108.53 a-g	MT64	10.86 a-k	MT33	51.84 a-j	MT29	2.70 a-h
108.08 b-g	MT37	10.84 a-l	MT35	51.44 a-j	MT36	2.67 a-1
107.48 b-h	<b>MT41</b>	10.83 a-l	MT32	51.30 a-j	MT42	2.58 a-j
107.23 b-1	MT42	10.81 a-l	MT50	51.19 a-k	<b>MT44</b>	2.44 b-k
106.93 b-j	MT18	10.70 a-l	MT47	50.89 a-k	<b>MT7</b>	2.41 b-m
106.48 с-1	MT52	10.68 a-m	МТ29	50.54 a-m	<b>MT6</b>	2.41 b-l
105.08 d-l	MT28	10.62 a-o	MT36	50.44 a-o	MT19	2.36 с-о
105.06 e-m	MT15	10.58 a-p	MT51	50.09 a-o	MT53	2.36 с-о
104.63 e-n	MT33	10.57 а-р	MT53	50.04 a-o	MT11	2.33 с-р
103.86 e-o	MT44	10.54 a-r	MT22	49.89 a-o	MT22	2.31 c-q
103.18 e-p	MT38	10.53 a-s	MT54	49.24 a-o	MT38	2.27 d-s
	PH (cm) 117.74 abc 100.52 f-u 92.86 n-A 105.80 c-k 119.56 a 118.96 ab 117.66 a-d 117.66 a-d 117.66 a-d 113.78 a-e 110.18 a-f 110.05 a-f 109.88 a-g 109.88 a-g 109.88 a-g 108.53 a-g 108.08 b-g 107.48 b-h 107.23 b-i 106.93 b-j 106.48 c-i 105.08 d-l 105.08 d-l 105.06 e-m 104.63 e-n 103.86 e-o 103.18 e-p	PH (cm)      Genotypes        117.74 abc      C1        100.52 f-u      C2        92.86 n-A      C3        105.80 c-k      C4        119.56 a      MT14        118.96 ab      MT35        117.66 a-d      MT31        117.66 a-d      MT31        117.66 a-d      MT31        117.66 a-d      MT31        117.66 a-d      MT48        113.78 a-e      MT36        110.18 a-f      MT27        110.05 a-f      MT17        109.88 a-g      MT40        108.53 a-g      MT40        108.53 a-g      MT41        107.23 b-1      MT42        106.93 b-j      MT18        106.48 c-1      MT28        105.08 d-l      MT28        105.06 e-m      MT15        104.63 e-n      MT33        103.86 e-o      MT44        103.18 e-p      MT38	PH (cm)      Genotypes      SL (cm)        117.74 abc      C1      10.78 a-1        100.52 f-u      C2      10.46 b-t        92.86 n-A      C3      10.51 a-s        105.80 c-k      C4      10.35 b-u        119.56 a      MT14      11.78 a        118.96 ab      MT35      11.45 ab        117.66 a-d      MT31      11.38 abc        117.66 a-d      MT48      11.24 a-d        113.78 a-e      MT36      11.20 a-e        110.18 a-f      MT27      11.07 a-g        110.05 a-f      MT17      10.96 a-h        109.88 a-g      MT40      10.86 a-l        108.53 a-g      MT64      10.86 a-l        107.23 b-1      MT42      10.81 a-l        107.23 b-1      MT42      10.81 a-l        106.93 b-j      MT18      10.70 a-l        106.48 c-1      MT52      10.68 a-m        105.08 d-l      MT28      10.62 a-o        105.06 e-m      MT15      10.58 a-p        104.63 e-n      MT33      10.57 a-p        103.86 e-o	PH (cm)      Genotypes      SL (cm)      Genotypes        117.74 abc      C1      10.78 a-1      C1        100.52 f-u      C2      10.46 b-t      C2        92.86 n-A      C3      10.51 a-s      C3        105.80 c-k      C4      10.35 b-u      C4        119.56 a      MT14      11.78 a      MT61        118.96 ab      MT35      11.45 ab      MT44        117.66 a-d      MT31      11.38 abc      MT30        117.66 a-d      MT48      11.24 a-d      MT31        113.78 a-e      MT36      11.20 a-e      MT59        110.18 a-f      MT27      11.07 a-g      MT28        110.05 a-f      MT17      10.96 a-h      MT27        109.88 a-g      MT40      10.86 a-l      MT42        108.08 b-g      MT37      10.84 a-l      MT35        107.23 b-1      MT41      10.83 a-l      MT32        107.23 b-1      MT42      10.68 a-m      MT29        105.08 d-l      MT28      10.62 a-o      MT36        106.48 c-1      MT52 <td>PH (cm)GenotypesSL (cm)GenotypesNGS (no)117.74 abcC110.78 a-1C142.98 e-x100.52 f-uC210.46 b-tC247.76 b-p92.86 n-AC310.51 a-sC348.56 a-o105.80 c-kC410.35 b-uC443.46 e-x119.56 aMT1411.78 aMT6159.84 a118.96 abMT3511.45 abMT4459.09 abc117.66 a-dMT3111.38 abcMT3056.44 a-d117.66 a-dMT3111.20 a-eMT5954.24 a-e110.18 a-fMT2711.07 a-gMT2854.04 a-f110.05 a-fMT1710.96 a-hMT2753.44 a-g109.88 a-gMT4010.86 a-lMT4252.19 a-1108.53 a-gMT6410.86 a-kMT3351.84 a-j107.48 b-hMT4110.83 a-lMT3251.30 a-j107.23 b-1MT4210.81 a-lMT5051.19 a-k106.48 c-1MT5210.68 a-mMT2950.54 a-m105.08 d-lMT2810.62 a-oMT3650.44 a-o105.08 d-lMT4210.81 a-lMT5051.19 a-k106.48 c-1MT5210.68 a-mMT2950.54 a-m105.08 d-lMT2810.62 a-oMT3650.44 a-o105.08 d-lMT5150.09 a-o104.63 e-nMT3310.57 a-pMT5350.04 a-o103.86 e-oMT4410.54 a-rMT2249.89 a-o103.18 e-p<!--</td--><td>PH (cm)      Genotypes      SL (cm)      Genotypes      NGS (no)      Genotypes        117.74 abc      C1      10.78 a-1      C1      42.98 e-x      C1        100.52 f-u      C2      10.46 b-t      C2      47.76 b-p      C2        92.86 n-A      C3      10.51 a-s      C3      48.56 a-o      C3        105.80 c-k      C4      10.35 b-u      C4      43.46 e-x      C4        119.56 a      MT14      11.78 a      MT61      59.84 a      MT14        118.96 ab      MT35      11.45 ab      MT44      59.09 abc      MT33        117.66 a-d      MT31      11.38 abc      MT30      56.44 a-d      MT28        117.66 a-d      MT48      11.20 a-e      MT59      54.24 a-e      MT30        110.18 a-f      MT27      11.07 a-g      MT27      53.44 a-g      MT32        110.05 a-f      MT17      10.96 a-h      MT27      53.44 a-g      MT32        109.88 a-g      MT40      10.86 a-l      MT42      52.19 a-1      MT36        107.48 b-h      MT41</td></td>	PH (cm)GenotypesSL (cm)GenotypesNGS (no)117.74 abcC110.78 a-1C142.98 e-x100.52 f-uC210.46 b-tC247.76 b-p92.86 n-AC310.51 a-sC348.56 a-o105.80 c-kC410.35 b-uC443.46 e-x119.56 aMT1411.78 aMT6159.84 a118.96 abMT3511.45 abMT4459.09 abc117.66 a-dMT3111.38 abcMT3056.44 a-d117.66 a-dMT3111.20 a-eMT5954.24 a-e110.18 a-fMT2711.07 a-gMT2854.04 a-f110.05 a-fMT1710.96 a-hMT2753.44 a-g109.88 a-gMT4010.86 a-lMT4252.19 a-1108.53 a-gMT6410.86 a-kMT3351.84 a-j107.48 b-hMT4110.83 a-lMT3251.30 a-j107.23 b-1MT4210.81 a-lMT5051.19 a-k106.48 c-1MT5210.68 a-mMT2950.54 a-m105.08 d-lMT2810.62 a-oMT3650.44 a-o105.08 d-lMT4210.81 a-lMT5051.19 a-k106.48 c-1MT5210.68 a-mMT2950.54 a-m105.08 d-lMT2810.62 a-oMT3650.44 a-o105.08 d-lMT5150.09 a-o104.63 e-nMT3310.57 a-pMT5350.04 a-o103.86 e-oMT4410.54 a-rMT2249.89 a-o103.18 e-p </td <td>PH (cm)      Genotypes      SL (cm)      Genotypes      NGS (no)      Genotypes        117.74 abc      C1      10.78 a-1      C1      42.98 e-x      C1        100.52 f-u      C2      10.46 b-t      C2      47.76 b-p      C2        92.86 n-A      C3      10.51 a-s      C3      48.56 a-o      C3        105.80 c-k      C4      10.35 b-u      C4      43.46 e-x      C4        119.56 a      MT14      11.78 a      MT61      59.84 a      MT14        118.96 ab      MT35      11.45 ab      MT44      59.09 abc      MT33        117.66 a-d      MT31      11.38 abc      MT30      56.44 a-d      MT28        117.66 a-d      MT48      11.20 a-e      MT59      54.24 a-e      MT30        110.18 a-f      MT27      11.07 a-g      MT27      53.44 a-g      MT32        110.05 a-f      MT17      10.96 a-h      MT27      53.44 a-g      MT32        109.88 a-g      MT40      10.86 a-l      MT42      52.19 a-1      MT36        107.48 b-h      MT41</td>	PH (cm)      Genotypes      SL (cm)      Genotypes      NGS (no)      Genotypes        117.74 abc      C1      10.78 a-1      C1      42.98 e-x      C1        100.52 f-u      C2      10.46 b-t      C2      47.76 b-p      C2        92.86 n-A      C3      10.51 a-s      C3      48.56 a-o      C3        105.80 c-k      C4      10.35 b-u      C4      43.46 e-x      C4        119.56 a      MT14      11.78 a      MT61      59.84 a      MT14        118.96 ab      MT35      11.45 ab      MT44      59.09 abc      MT33        117.66 a-d      MT31      11.38 abc      MT30      56.44 a-d      MT28        117.66 a-d      MT48      11.20 a-e      MT59      54.24 a-e      MT30        110.18 a-f      MT27      11.07 a-g      MT27      53.44 a-g      MT32        110.05 a-f      MT17      10.96 a-h      MT27      53.44 a-g      MT32        109.88 a-g      MT40      10.86 a-l      MT42      52.19 a-1      MT36        107.48 b-h      MT41

Table 2. Mean performance of checks and mutant bread wheat lines for PH, SL, NGS and GWS

MT58	103.16 e-r	MT34	10.50 a-s	MT48	49.19 a-o	MT15	2.27 d-r
MT26	103.03 e-r	MT59	10.50 a-s	MT15	48.39 a-p	MT51	2.27 d-r
MT7	102.98 e-r	MT16	10.35 b-u	MT58	47.54 b-p	MT24	2.24 e-s
MT48	102.03 e-s	MT30	10.33 b-u	MT45	47.29 b-p	MT46	2.23 f-t
MT53	101.86 e-s	MT23	10.31 b-u	MT57	46.84 c-r	MT23	2.19 g-t
MT60	101.76 e-s	MT22	10.27 b-u	<b>MT7</b>	46.44 d-t	MT10	2.17 g-t
MT57	100.96 f-t	MT29	10.22 b-u	MT24	46.19 d-t	MT20	2.15 g-t
MT41	99.23 f-v	MT54	10.19 b-u	MT2	45.94 d-u	MT47	2.15 g-t
MT24	98.13 f-v	MT10	10.11 c-v	MT20	45.09 d-v	MT48	2.15 g-t
MT62	97.76 f-y	MT21	10.04d-v	MT10	45.04 d-v	MT34	2.13 g-t
MT31	97.71 f-y	MT62	10.02 d-w	MT37	44.74 d-v	MT40	2.12 g-t
MT42	97.63 f-y	MT13	10.00 d-x	MT19	44.59 d-v	MT52	2.10 g-t
MT23	97.33 g-у	MT47	9.96 d-x	MT13	44.54 d-v	MT21	2.09 g-t
MT47	97.33 g-у	MT56	9.94 d-x	MT34	44.54 d-v	MT54	2.09 g-t
MT2	95.58 h-z	MT45	9.86 e-x	MT46	44.49 d-w	MT25	2.07 h-t
MT32	94.91 h-z	MT26	9.81 f-x	MT62	44.44 d-w	MT58	2.05 j-u
MT46	94.73 1-z	MT20	9.80 f-x	MT56	44.34 d-w	MT17	2.05 1-u
MT33	94.61 1-z	MT2	9.78 g-x	MT63	44.34 d-w	MT2	2.03 j-u
MT44	94.33 k-z	MT6	9.78 g-x	MT17	44.19 d-w	MT16	2.03 j-u
MT19	94.13 k-z	MT60	9.74 g-y	MT41	43.59 e-x	MT1	2.02 j-u
MT8	93.31 l-z	MT11	9.73 g-у	MT23	43.29 e-x	МТ9	2.01 j-u
MT1	93.08 m-A	МТ9	9.69 h-y	MT11	42.34 e-y	MT59	2.00 j-u
MT49	92.73 o-A	MT61	9.69 h-y	MT16	41.49 f-y	MT61	1.99 j <b>-</b> u
MT43	92.53 o-A	MT1	9.64 1 <b>-</b> y	MT40	41.29 g-у	MT13	1.98 j-u
МТ39	92.30 o-A	MT46	9.62 ј-у	MT1	41.14 g-y	MT50	1.98 j-u
MT28	91.90 o-A	MT55	9.60 k-y	MT38	41.14 g-y	MT4	1.97 j-u
MT37	91.30 o-A	MT65	9.58 l-y	MT55	40.84 h-y	MT41	1.96 k-u
MT65	91.25 p-A	MT19	9.45 m-z	MT60	39.74 1-у	MT57	1.96 k-u
MT20	90.83 r-A	MT12	9.32 n-z	MT52	39.39 ј-у	MT26	1.95 k-u
МТ30	90.70 r-A	MT63	9.30 o-A	MT4	38.54 k-y	MT5	1.92 k-u
MT18	90.63 r-A	MT51	9.28 p-A	MT64	38.24 l-y	MT18	1.88 k-u
MT14	90.53 r-A	MT7	9.26 q-A	MT21	37.99 m-y	MT65	1.88 k-u
MT16	89.53 s-A	MT43	9.26 q-A	МТ9	37.94 n-y	МТ39	1.86 k-u
MT21	89.53 s-A	MT57	9.22 r-A	MT18	37.89 о-у	MT62	1.85 k-u
MT50	89.03 t-A	MT50	9.16 s-A	MT3	36.14 p-y	MT43	1.84 l-u
MT27	88.70 t-A	MT8	9.15 t-A	MT6	34.94 q-y	MT45	1.84 l-u
MT38	88.50 t-A	MT32	9.06 u-A	MT12	34.74 q-y	MT56	1.79 m-u
MT15	88.33 t-A	MT39	8.80 v-B	MT26	34.49 r-y	MT12	1.74 o-u
MT36	87.51 u-A	MT24	8.74 w-B	MT65	34.14 s-y	MT63	1.72 p-u
MT29	87.41 u-A	MT58	8.69 x-B	MT25	33.99 t-y	MT49	1.70 r-u
MT22	86./3 v-A	MT49	8.43 y-B	MT43	33.39 u-y	MT60	1.70 r-u
MT35	86.20 v-A	MT53	8.24 zAB	MT39	33.04 v-y	MT64	1./0 r-u
MT52	83.03 yzA	MT5	8.18 zAB	MT5	31.94 wxy	MT3	1.63 stu
MT34	80.40 AB	MT3	8.08 AB	MT8	31.44 xy	MT55	1.59 tu
MT17	69.13 B	MT4	7.61 B	MT49	30.59 y	MT8	1.43 u

The identical letters indicate statistical groups of identical values with 0.01 confidence level by the Student-Newman-Keul Test (SNKT)

important selection criteria in wheat breeding as well. Previous studies have shown that mutagen applications cause significant changes in spike length in wheat (Mohammad et al., 2004; Farag & El-Khawaga, 2013). In the study, the spike length of 65 advanced bread wheat mutant lines ranged from 7.61 to 11.78 cm, and the spike length of check genotypes ranged from 10.46 to 10.78 cm (Table 2). Among the mutant lines examined in the experiment, 13 lines (MT14, 35, 31, 48, 36, 27, 17, 25, 64, 40, 37, 41 and 42) formed longer spike than all check genotypes. These results are similar to those of Githinji & Birithia (2015), who also reported highly significant differences for spike length and mutant bread wheat lines produced the longest spike as compared to the parent wheat variety.

The number of grains per spike (NGS) (no): In the case of wheat improvement, the number of grains per spike is one of the selection criteria considered as the most important main yield component in wheat breeding studies. It has been determined that 65 advanced bread wheat mutant lines tested in the M6 genome exhibit a wide variation in the number of seeds per spike from 30.59 to 59.88, while it is as low as 42.98 to 47.76 no (Table 2). Among the tested lines, 21 lines (MT61, 44, 30, 31, 59, 28, 27, 14, 42, 33, 35, 32, 50, 47, 29, 36, 51, 53, 22, 54 and 48) was found to have a higher number of grains per spike than the check IBWSN-4 genotype, giving the highest mean and other parents. These results are similar to the results of Anter (2021), who explained that 22% of the mutant lines have more grains per spike than the parents.

Grain weight per spike (GWS) (g): One of the most important yielding factors affecting wheat yield is the grain weight per spike and it is accepted as one of the most important selection criteria in wheat breeding as well. In terms of this character, means of the grain weight per spike for the 65 advanced bread wheat mutant lines in the M6 generation ranged from 1.43 to 3.13 g, while the check genotypes ranged from 1.95 to 2.30 g indicating variation was wider in lines than that of check genotypes (Table 2). Among them, 19 mutant lines (MT14, 31, 28, 33, 30, 27, 32, 35, 37, 29, 36, 42, 44, 6, 7, 53, 19, 11 and 22) were a higher grain weight per spike than the IBWSN-4 check genotype, which gave the highest averages. These results suggest that these 19 mutant lines should be considered as a priority for grain weight per spike and agree with the results of Sari et al. (2016) who have reported that 32% of the mutant lines have more grain weight per spike than the parents.

Harvest index (HI) (%): The harvest index, which is the ratio of economic yield to biological yield, is an important selection criterion. It is desirable that the harvest index be as high as possible (even closer to 50%) in wheat breeding. The harvest index values of sixty-five advanced bread wheat mutant lines ranged between 32.57% and 48.37%, and between 35.39% and 40.60% in check genotypes. 35 mutant lines (MT33, 37, 30, 47, 11, 29, 42, 50, 44, 48, 46, 27, 32, 28, 21, 35, 31, 51, 4, 19, 17, 20, 7, 6, 14, 52, 39, 15, 10, 62, 58, 36, 22, 43 and 9) were the highest harvest index value compared to check genotypes (Table 3). Fourteen mutant lines (MT33, 37, 30, 47, 11, 29, 42, 50, 44, 48, 46, 27, 32 and 28) performed well with harvest index averages of over 45% and the currently accepted harvest index value is close to 50%. The findings of Rahimi & Bahrani (2011), which explains that lines with higher harvest index averages than the germplasm can be developed with gamma irradiation, proves our results.

Thousand grain weight (TGW) (g): Thousand grain weight (g) or seed index which is an indicator of grain size and flour yield in wheat is an important main yield contributing trait. In our study, the thousand grain weight means ranged from 34.13 to 55.93 g for the 65 advanced bread wheat mutant lines ranged from 36.9 to 47.04 g for the check genotypes. Among the bread wheat mutant lines, 30 lines (MT43, 41, 5, 49, 21, 46, 65, 39, 8, 7, 20, 29, 52, 40, 19, 31, 28, 30, 36, 38, 6, 23, 24, 18, 9, 37, 12, 35, 25 and 53) were a higher 1000-grain weight than the check genotype. These results show that a wide variation can be achieved with the application of mutagen for 1000 grain weights. Fifteen mutant lines (MT43, 41, 5, 49, 21, 46, 65, 39, 8, 7, 20, 29, 52, 40 and 19)

draw attention with an average of thousand grain weight above 50 g (Table 3). Our results are similar to the results of Singh & Balyan (2009) who illustrated mutant lines with larger grains than the parents can be obtained as a result of gamma irradiation, while it is contradictory to the findings by Öztürk et al. (2020) who the mutant lines have a lower thousand-grain weight than their parents.

*Grain yield (GY) (kg ha<sup>-1</sup>):* The main purpose of plant improvement is to increase grain yield obtained from the unit area. Grain yield is polygenic

Table 3. Mean	performance of	of checks	and mutant	bread v	wheat l	ines for	r HI.	TGW	and (	GY
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Genotypes	HI (%)	Genotypes	TGW (g)	Genotypes	GY (kg ha <sup>-1</sup> )
C1	35.39 o-t	C1	36.9 w-z	C1	6074 p-B
C2	40.60 a-t	C2	42.78 n-x	C2	8556 a-h
C3	40.50 a-t	C3	44.94 1-v	C3	8362 a-j
C4	38.00 g-t	C4	47.04 c-s	C4	6836 k-y
MT33	48.37 a	MT43	55.93 a	MT53	9515 a
MT37	48.28 abc	MT41	55.53 ab	MT19	9412 ab
MT30	48.16 abc	MT5	53.48 a-d	MT33	9232 abc
MT47	47.82 a-d	MT49	53.33 a-d	MT35	9122 a-d
MT11	47.53 a-e	MT21	52.43 a-1	<b>MT14</b>	9112 a-e
MT29	46.26 a-f	MT46	52.13 a-j	MT20	9082 a-e
MT42	46.18 a-f	MT65	52.03 a-j	<b>MT42</b>	8840 a-f
MT50	46.12 a-f	МТ39	52.01 a-j	<b>MT40</b>	8820 a-g
MT44	45.90 a-f	<b>MT8</b>	51.68 a-j	<b>MT27</b>	8632 a-h
MT48	45.90 a-f	MT7	50.88 a-k	MT15	8542 a-1
MT46	45.85 a-f	MT20	50.53 a-l	<b>MT44</b>	8250 a-k
MT27	45.81 a-g	MT29	50.51 a-l	MT32	7982 a-l
MT32	45.67 a-g	MT52	50.33 a-1	MT29	7942 a-n
MT28	45.21 a-h	MT40	50.13 a-l	<b>MT16</b>	7942 a-m
MT21	44.93 a-1	MT19	50.03 a-m	MT28	7792 b-о
MT35	44.54 a-l	MT31	49.91 a-m	MT11	7702 с-о
MT31	44.49 a-l	MT28	49.41 a-n	MT43	7590 с-о
MT51	44.41 a-l	MT30	49.41 a-n	MT58	7585 с-р
MT4	44.23 a-l	MT36	49.41 a-n	MT12	7572 с-р
MT19	43.92 a-m	MT38	49.31 a-n	MT57	7495 d-q
MT17	43.86 a-m	MT6	48.88 b-o	MT17	7462 e-r
MT20	43.57 a-n	MT23	48.73 b-p	MT13	7392 f-r
MT7	43.47 a-o	MT24	48.73 b-p	MT31	7272 g-s
MT6	42.96 a-p	MT18	48.53 c-p	MT7	7172 g-t
MT14	42.37 a-q	МТ9	48.38 c-p	MT1	7152 g-t
MT52	42.17 a-q	MT37	47.91 c-q	MT56	7145 h-t
МТ39	42.16 a-q	MT12	47.48 c-r	MT30	7142 h-u
MT15	42.05 a-q	MT32	47.31 c-r	MT37	7122 i-u
MT10	42.02 a-r	MT25	47.13 c-s	MT38	7102 i-v
MT62	41.97 a-r	MT53	47.13 c-s	MT34	7072 j-v
MT58	41.60 a-r	MT26	47.03 c-s	MT55	7025 j-w
МТ36	41.00 a-s	MT51	47.03 c-s	MT6	6952 j-x
MT22	40.94 a-s	MT22	46.93 c-s	MT22	6862 k-y
MT43	40.83 a-s	MT11	46.38 d-u	MT21	6822 k-y
МТ9	40.62 a-t	MT14	46.33 d-u	MT52	6730 k-y
MT65	40.19 a-t	MT47	46.03 e-u	MT3	6642 k-y
MT2	40.15 b-t	MT34	46.01 e-u	MT54	6565 k-z
MT38	40.10 c-t	MT3	45.88 e-u	MT18	6482 l-A
MT24	39.75 d-t	МТ33	45.71 e-v	MT23	6482 l-A
МТ60	39.65 d-t	MT4	45.68 e-v	MT36	6442 m-A

MT1	39.53 e-t	MT48	45.43 f-v	MT26	6412 m-A
MT53	39.47 e-t	MT10	45.38 g-v	MT59	6275 m-B
MT57	39.42 e-t	MT45	45.33 h-v	<b>MT10</b>	6262 m-B
MT49	39.16 f-t	MT1	44.98 h-v	MT4	6222 о-В
MT18	38.94 f-t	MT42	44.83 j-v	MT5	6202 о-В
MT59	38.81 f-t	MT15	44.73 k-v	MT51	6170 о-В
MT13	38.68 f-t	MT27	44.71 k-v	<b>MT41</b>	6160 о-В
MT12	38.67 f-t	MT63	44.53 k-v	MT46	6130 о-В
MT54	38.61 f-t	MT13	44.28 l-v	MT49	6120 о-В
MT8	38.26 f-t	MT58	43.93 l-v	<b>MT47</b>	5960 p-B
MT41	37.81 g-t	<b>MT44</b>	43.43 m-w	MT64	5865 q-B
MT56	37.29 h-t	MT35	43.21 m-x	<b>MT9</b>	5862 q-B
MT5	36.94 1-t	MT16	42.63 n-x	MT65	5855 q-B
MT16	36.94 1-t	MT57	42.03 о-у	<b>MT8</b>	5852 q-В
MT45	36.58 k-t	MT2	41.18 р-у	MT48	5800 r-B
MT40	36.53 k-t	MT50	40.33 q-z	MT60	5565 s-B
MT26	36.50 l-t	MT56	39.93 s-z	MT61	5525 t-B
MT23	35.79 m-t	MT64	39.73 t-z	MT62	5455 u-B
MT3	35.61 n-t	MT54	39.63 u-z	<b>MT24</b>	5422 v-B
MT55	35.45 n-t	MT62	38.93 v-z	MT2	5362 w-B
MT34	35.17 p-t	MT55	37.03 w-z	MT45	5290 x-B
MT61	35.02 p-t	MT60	36.73 w-z	MT39	5252 у-В
MT63	34.38 q-t	MT17	36.43 xyz	MT50	4960 zAB
MT25	33.89 rst	MT61	34.23z	MT63	4875 AB
MT64	32.57 t	MT59	34.13 z	MT25	4712 B

The identical letters indicate statistical groups of identical values with 0.01 confidence level by the Student-Newman-Keul Test (SNKT)

and is highly influenced by cultivar, environment and cultivar x environment interactions. The average performance of grain yield (kg ha<sup>-1</sup>) of all genotypes is depicted in (Table 3). In the study, the grain yield of advanced bread wheat mutant lines ranged from 4712-9415 kg ha<sup>-1</sup>, while the grain yield of check genotypes ranged from 6074-8556 kg ha (Table 3). Considering the other characteristics examined, it is seen that there is a higher variation in grain yield between mutant genotypes obtained by mutagen application. Results regarding the trait revealed that a higher grain yield was observed in 9 mutant lines (MT53, 19, 33, 35, 14, 20, 42, 40 and 27) as compared to the other genotypes. Ayub et al. (1989) stated that the effect of different doses of gamma rays irradiation on the grain yield of wheat varieties responded differently in different varieties. Our results are in confirmation with those of Oztürk et al. (2020) who explained that 33% of the

mutant lines have more grain yield than the parents and check genotypes. As a result, it can be said that it would be appropriate to take these 9 mutant lines into the pre-registration yield trials in terms of grain yield.

# CONCLUSION

Regarding plant height, among the 65 mutant lines, 13 mutant lines below 90 cm are recommended for our region to avoid lodging problems. Thirteen mutant lines gave a long spike than 10.5 cm and 1 mutant line gave a spike longer than 11.0 cm. In total, 21 lines were higher the number of grains per spike, which is one of the important yield criteria than the parents, and of them14 mutants gave 50 grains, 3 mutant lines gave more than 55 grains. When grain weight per spike values of mutant lines are examined, it is shown that 19 mutant lines were a higher grain weight per spike than the parent which gave the highest averages, among them 9 mutant lines were more than 2.5 g, 2 mutant genotypes were above 3.0 g. Fourteen mutant lines gave well with harvest index averages of over 45% and the currently accepted harvest index value is close to 50%. Among the advanced bread wheat mutant lines, 30 lines were a higher 1000-grain weight than the parent genotypes. The higher grain yield was recorded in 9 mutant lines as compared to the other mutant genotypes and their parents, of the 4 mutant lines yielded higher than 850 kg ha<sup>-1</sup> and 6 lines vielded more than 900 kg ha-1. The mutant MT53, MT19, MT33, MT35, MT14, MT20, MT42, MT40 and MT27 were out yielded than the other mutant lines and parent varieties. Among them, MT35, MT14 and MT27 were over-performed for all yield components except thousand grain weight. As a result, MT35, MT14, MT27, MT19, MT20 and MT15 should be included in further breeding programs for exploitation of its genetic potential in new cross combinations. The fact that the majority of selected and promising mutants originated from populations irradiated with 100 and 200 Gy gamma rays (Table 1), suggests that these gamma doses may be more beneficial for yield increase in wheat mutation breeding.

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